Full wwPDB NMR Structure Validation Report (i)

May 29, 2020 – 04:40 am BST

PDB ID	:	2N0Q
Title	:	N2-dG-IQ modified DNA at the G1 position of the NarI recognition sequence
Authors	:	Stavros, K.; Hawkins, E.; Rizzo, C.; Stone, M.
Deposited on	:	2015-03-11

This is a Full wwPDB NMR Structure Validation Report for a publicly released PDB entry.

We welcome your comments at *validation@mail.wwpdb.org*
A user guide is available at
https://www.wwpdb.org/validation/2017/NMRValidationReportHelp
with specific help available everywhere you see the (i) symbol.

The following versions of software and data (see references (1)) were used in the production of this report:

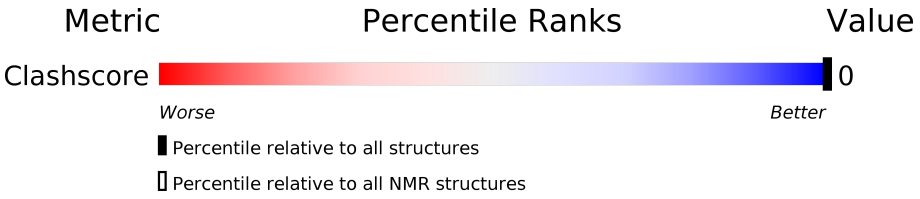
Cyrange	:	Kirchner and Güntert (2011)
NmrClust	:	Kelley et al. (1996)
MolProbity	:	4.02b-467
Mogul	:	1.8.5 (274361), CSD as541be (2020)
Percentile statistics	:	20191225.v01 (using entries in the PDB archive December 25th 2019)
RCI	:	v_1n_11_5_13_A (Berjanski et al., 2005)
PANAV	:	Wang et al. (2010)
ShiftChecker	:	2.11
Ideal geometry (proteins)	:	Engh & Huber (2001)
Ideal geometry (DNA, RNA)	:	Parkinson et al. (1996)
Validation Pipeline (wwPDB-VP)	:	2.11

1 Overall quality at a glance

The following experimental techniques were used to determine the structure:
SOLUTION NMR

The overall completeness of chemical shifts assignment is 29%.

Percentile scores (ranging between 0-100) for global validation metrics of the entry are shown in the following graphic. The table shows the number of entries on which the scores are based.



Metric	Whole archive (#Entries)	NMR archive (#Entries)
Clashscore	158937	12864

The table below summarises the geometric issues observed across the polymeric chains and their fit to the experimental data. The red, orange, yellow and green segments indicate the fraction of residues that contain outliers for >=3, 2, 1 and 0 types of geometric quality criteria. A cyan segment indicates the fraction of residues that are not part of the well-defined cores, and a grey segment represents the fraction of residues that are not modelled. The numeric value for each fraction is indicated below the corresponding segment, with a dot representing fractions <=5%

Mol	Chain	Length	Quality of chain
1	A	12	8% 92%
2	B	12	83% 17%

2 Ensemble composition and analysis

This entry contains 10 models. This entry does not contain polypeptide chains, therefore identification of well-defined residues and clustering analysis are not possible. All residues are included in the validation scores.

3 Entry composition (i)

There are 2 unique types of molecules in this entry. The entry contains 780 atoms, of which 279 are hydrogens and 0 are deuteriums.

- Molecule 1 is a DNA chain called DNA_(5'-D(*CP*TP*CP*(IQG)P*GP*CP*GP*CP*CP *AP*TP*C)-3').

Mol	Chain	Residues	Atoms						Trace
			Total	C	H	N	O	P	
1	A	12	397	125	144	46	71	11	0

- Molecule 2 is a DNA chain called DNA_(5'-D(*GP*AP*TP*GP*GP*CP*GP*CP*CP*GP *AP*G)-3').

Mol	Chain	Residues	Atoms						Trace
			Total	C	H	N	O	P	
2	B	12	383	117	135	51	69	11	0

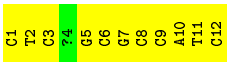
4 Residue-property plots

4.1 Average score per residue in the NMR ensemble

These plots are provided for all protein, RNA and DNA chains in the entry. The first graphic is the same as shown in the summary in section 1 of this report. The second graphic shows the sequence where residues are colour-coded according to the number of geometric quality criteria for which they contain at least one outlier: green = 0, yellow = 1, orange = 2 and red = 3 or more. Stretches of 2 or more consecutive residues without any outliers are shown as green connectors. Residues which are classified as ill-defined in the NMR ensemble, are shown in cyan with an underline colour-coded according to the previous scheme. Residues which were present in the experimental sample, but not modelled in the final structure are shown in grey.

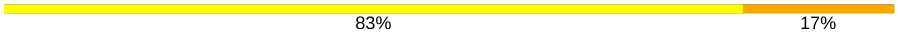
• Molecule 1: DNA_(5'-D(*CP*TP*CP*(IQG)P*GP*CP*GP*CP*CP*AP*TP*C)-3')

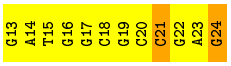
Chain A:



• Molecule 2: DNA_(5'-D(*GP*AP*TP*GP*GP*CP*GP*CP*CP*GP*AP*G)-3')

Chain B:





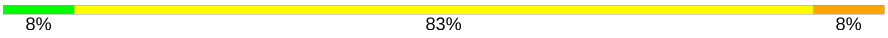
4.2 Scores per residue for each member of the ensemble

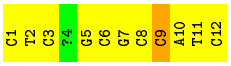
Colouring as in section 4.1 above.

4.2.1 Score per residue for model 1

• Molecule 1: DNA_(5'-D(*CP*TP*CP*(IQG)P*GP*CP*GP*CP*CP*AP*TP*C)-3')

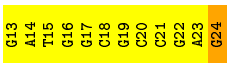
Chain A:





• Molecule 2: DNA_(5'-D(*GP*AP*TP*GP*GP*CP*GP*CP*CP*GP*AP*G)-3')

Chain B:

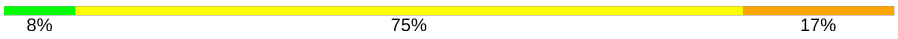


4.2.2 Score per residue for model 2

• Molecule 1: DNA_(5'-D(*CP*TP*CP*(IQG)P*GP*CP*GP*CP*CP*AP*TP*C)-3')

Chain A: 

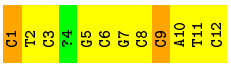
• Molecule 2: DNA_(5'-D(*GP*AP*TP*GP*GP*CP*GP*CP*CP*GP*AP*G)-3')

Chain B: 

4.2.3 Score per residue for model 3

• Molecule 1: DNA_(5'-D(*CP*TP*CP*(IQG)P*GP*CP*GP*CP*CP*AP*TP*C)-3')

Chain A: 



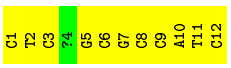
• Molecule 2: DNA_(5'-D(*GP*AP*TP*GP*GP*CP*GP*CP*CP*GP*AP*G)-3')

Chain B: 

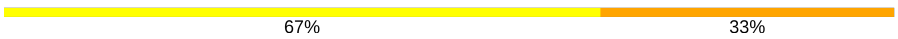
4.2.4 Score per residue for model 4

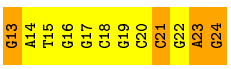
• Molecule 1: DNA_(5'-D(*CP*TP*CP*(IQG)P*GP*CP*GP*CP*CP*AP*TP*C)-3')

Chain A: 



• Molecule 2: DNA_(5'-D(*GP*AP*TP*GP*GP*CP*GP*CP*CP*GP*AP*G)-3')

Chain B: 

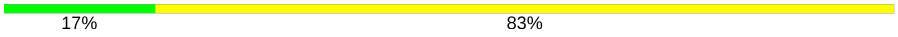


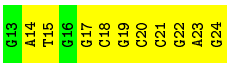
4.2.5 Score per residue for model 5

• Molecule 1: DNA_(5'-D(*CP*TP*CP*(IQG)P*GP*CP*GP*CP*CP*AP*TP*C)-3')

Chain A: 

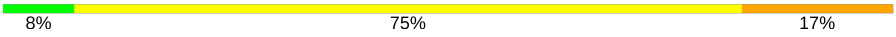
• Molecule 2: DNA_(5'-D(*GP*AP*TP*GP*GP*CP*GP*CP*CP*GP*AP*G)-3')

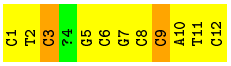
Chain B: 



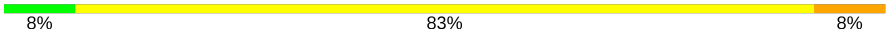
4.2.6 Score per residue for model 6

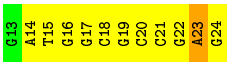
• Molecule 1: DNA_(5'-D(*CP*TP*CP*(IQG)P*GP*CP*GP*CP*CP*AP*TP*C)-3')

Chain A: 



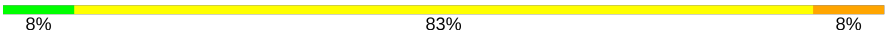
• Molecule 2: DNA_(5'-D(*GP*AP*TP*GP*GP*CP*GP*CP*CP*GP*AP*G)-3')

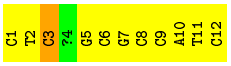
Chain B: 



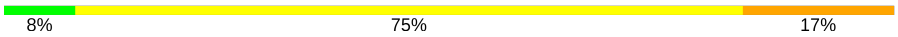
4.2.7 Score per residue for model 7

• Molecule 1: DNA_(5'-D(*CP*TP*CP*(IQG)P*GP*CP*GP*CP*CP*AP*TP*C)-3')

Chain A: 



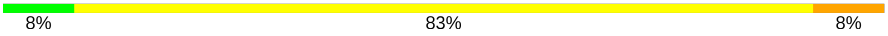
• Molecule 2: DNA_(5'-D(*GP*AP*TP*GP*GP*CP*GP*CP*CP*GP*AP*G)-3')

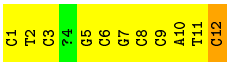
Chain B: 

4.2.8 Score per residue for model 8

• Molecule 1: DNA_(5'-D(*CP*TP*CP*(IQG)P*GP*CP*GP*CP*CP*AP*TP*C)-3')

Chain A:





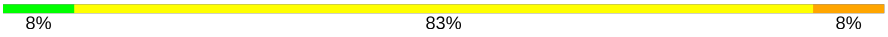
• Molecule 2: DNA_(5'-D(*GP*AP*TP*GP*GP*CP*GP*CP*CP*GP*AP*G)-3')

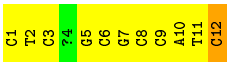
Chain B:

4.2.9 Score per residue for model 9

• Molecule 1: DNA_(5'-D(*CP*TP*CP*(IQG)P*GP*CP*GP*CP*CP*AP*TP*C)-3')

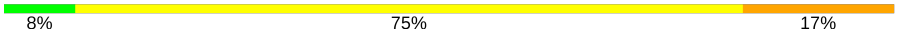
Chain A:





• Molecule 2: DNA_(5'-D(*GP*AP*TP*GP*GP*CP*GP*CP*CP*GP*AP*G)-3')

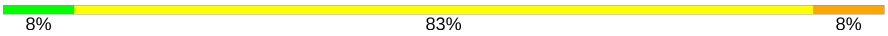
Chain B:

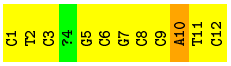


4.2.10 Score per residue for model 10

• Molecule 1: DNA_(5'-D(*CP*TP*CP*(IQG)P*GP*CP*GP*CP*CP*AP*TP*C)-3')

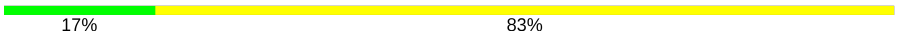
Chain A:





• Molecule 2: DNA_(5'-D(*GP*AP*TP*GP*GP*CP*GP*CP*CP*GP*AP*G)-3')

Chain B:



5 Refinement protocol and experimental data overview

The models were refined using the following method: *simulated annealing*.

Of the 10 calculated structures, 10 were deposited, based on the following criterion: *structures with the lowest energy*.

The following table shows the software used for structure solution, optimisation and refinement.

Software name	Classification	Version
AMBER	refinement	
AMBER	structure solution	

The following table shows chemical shift validation statistics as aggregates over all chemical shift files. Detailed validation can be found in section 6 of this report.

Chemical shift file(s)	input_cs.cif
Number of chemical shift lists	1
Total number of shifts	150
Number of shifts mapped to atoms	146
Number of unparsed shifts	2
Number of shifts with mapping errors	0
Number of shifts with mapping warnings	2
Assignment completeness (well-defined parts)	29%

No validations of the models with respect to experimental NMR restraints is performed at this time.

COVALENT-GEOMETRY INFOmissingINFO

5.1 Too-close contacts

In the following table, the Non-H and H(model) columns list the number of non-hydrogen atoms and hydrogen atoms in each chain respectively. The H(added) column lists the number of hydrogen atoms added and optimized by MolProbity. The Clashes column lists the number of clashes averaged over the ensemble.

Mol	Chain	Non-H	H(model)	H(added)	Clashes
All	All	5010	2790	2790	-

The all-atom clashscore is defined as the number of clashes found per 1000 atoms (including hydrogen atoms). The all-atom clashscore for this structure is -.

There are no clashes.

5.2 Torsion angles

5.2.1 Protein backbone

There are no protein molecules in this entry.

5.2.2 Protein sidechains

There are no protein molecules in this entry.

5.2.3 RNA

There are no RNA molecules in this entry.

5.3 Non-standard residues in protein, DNA, RNA chains

There are no non-standard protein/DNA/RNA residues in this entry.

5.4 Carbohydrates

There are no carbohydrates in this entry.

5.5 Ligand geometry

There are no ligands in this entry.

5.6 Other polymers

There are no such molecules in this entry.

5.7 Polymer linkage issues

There are no chain breaks in this entry.

6 Chemical shift validation

The completeness of assignment taking into account all chemical shift lists is 29% for the well-defined parts and 29% for the entire structure.

6.1 Chemical shift list 1

File name: input_cs.cif

Chemical shift list name: *assigned_chem_shift_list_1*

6.1.1 Bookkeeping

The following table shows the results of parsing the chemical shift list and reports the number of nuclei with statistically unusual chemical shifts.

Total number of shifts	150
Number of shifts mapped to atoms	146
Number of unparsed shifts	2
Number of shifts with mapping errors	0
Number of shifts with mapping warnings	2
Number of shift outliers (ShiftChecker)	0

The following errors were found when reading this chemical shift list.

- Chemical shift has been reported more than once. All 2 occurences are reported below.

Shift ID	Chain	Res	Type	Atom	Shift Data		
					Value	Uncertainty	Ambiguity
34	A	4	IQG	H3'	5.122	?	.
35	A	4	IQG	H8	8.302	?	.

The following assigned chemical shifts were not mapped to the molecules present in the coordinate file.

- No matching atoms found in structure. All 2 occurences are reported below.

Chain	Res	Type	Atom	Shift Data		
				Value	Uncertainty	Ambiguity
A	4	IQG	H2"	2.77	-1.0	1
A	4	IQG	H2'	2.669	-1.0	1

6.1.2 Chemical shift referencing (i)

No chemical shift referencing corrections were calculated (not enough data).

6.1.3 Completeness of resonance assignments (i)

The following table shows the completeness of the chemical shift assignments for the well-defined regions of the structure. The overall completeness is 29%, i.e. 133 atoms were assigned a chemical shift out of a possible 462. 0 out of 0 assigned methyl groups (LEU and VAL) were assigned stereospecifically.

	Total	^{1}H	^{13}C	^{15}N
Backbone	0/0 (—%)	0/0 (—%)	0/0 (—%)	0/0 (—%)
Sidechain	0/0 (—%)	0/0 (—%)	0/0 (—%)	0/0 (—%)
Aromatic	0/0 (—%)	0/0 (—%)	0/0 (—%)	0/0 (—%)
Overall	133/462 (29%)	133/278 (48%)	0/153 (0%)	0/31 (0%)

The following table shows the completeness of the chemical shift assignments for the full structure. The overall completeness is 29%, i.e. 133 atoms were assigned a chemical shift out of a possible 462. 0 out of 0 assigned methyl groups (LEU and VAL) were assigned stereospecifically.

	Total	^{1}H	^{13}C	^{15}N
Backbone	0/0 (—%)	0/0 (—%)	0/0 (—%)	0/0 (—%)
Sidechain	0/0 (—%)	0/0 (—%)	0/0 (—%)	0/0 (—%)
Aromatic	0/0 (—%)	0/0 (—%)	0/0 (—%)	0/0 (—%)
Overall	133/462 (29%)	133/278 (48%)	0/153 (0%)	0/31 (0%)

6.1.4 Statistically unusual chemical shifts (i)

There are no statistically unusual chemical shifts.

6.1.5 Random Coil Index (RCI) plots (i)

No *random coil index* (RCI) plot could be generated from the current chemical shift list (assigned_chem_shift_list_1). RCI is only applicable to proteins.