



Full wwPDB NMR Structure Validation Report ⓘ

May 31, 2020 – 11:19 am BST

PDB ID : 6MCE
Title : Solution structure of HIV-1 TAR with Tat RNA Binding Domain
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Deposited on : 2018-08-31

This is a Full wwPDB NMR Structure Validation Report for a publicly released PDB entry.

We welcome your comments at validation@mail.wwpdb.org

A user guide is available at

<https://www.wwpdb.org/validation/2017/NMRValidationReportHelp>

with specific help available everywhere you see the ⓘ symbol.

The following versions of software and data (see [references ⓘ](#)) were used in the production of this report:

Cyrange : Kirchner and Güntert (2011)
NmrClust : Kelley et al. (1996)
MolProbity : 4.02b-467
Percentile statistics : 20191225.v01 (using entries in the PDB archive December 25th 2019)
RCI : v_1n_11_5_13_A (Berjanski et al., 2005)
PANAV : Wang et al. (2010)
ShiftChecker : 2.11
Ideal geometry (proteins) : Engh & Huber (2001)
Ideal geometry (DNA, RNA) : Parkinson et al. (1996)
Validation Pipeline (wwPDB-VP) : 2.11

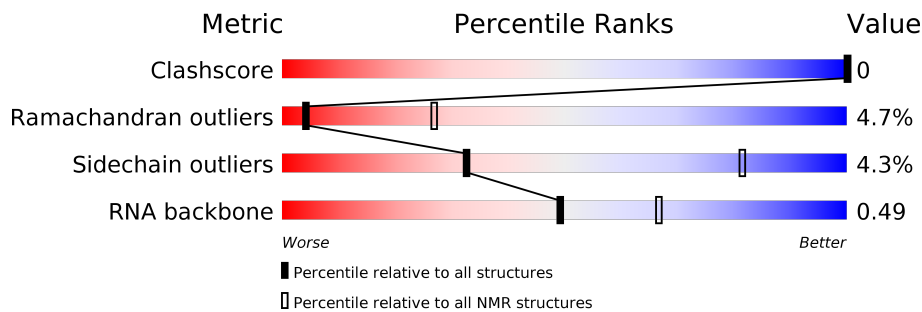
1 Overall quality at a glance

The following experimental techniques were used to determine the structure:

SOLUTION NMR

The overall completeness of chemical shifts assignment is 51%.

Percentile scores (ranging between 0-100) for global validation metrics of the entry are shown in the following graphic. The table shows the number of entries on which the scores are based.



Metric	Whole archive (#Entries)	NMR archive (#Entries)
Clashscore	158937	12864
Ramachandran outliers	154571	11451
Sidechain outliers	154315	11428
RNA backbone	4643	676

The table below summarises the geometric issues observed across the polymeric chains and their fit to the experimental data. The red, orange, yellow and green segments indicate the fraction of residues that contain outliers for ≥ 3 , 2, 1 and 0 types of geometric quality criteria. A cyan segment indicates the fraction of residues that are not part of the well-defined cores, and a grey segment represents the fraction of residues that are not modelled. The numeric value for each fraction is indicated below the corresponding segment, with a dot representing fractions $\leq 5\%$

Mol	Chain	Length	Quality of chain
1	A	30	 23% 63% 13%
2	B	17	 76% 18% 6%

2 Ensemble composition and analysis ⓘ

This entry contains 10 models.

Cyrange was unable to find well-defined residues.

Error message: Only domains with < 8 residues could be identified.

NmrClust was unable to cluster the ensemble.

Error message: Wrapper check: not enough residues in core to run NmrClust

3 Entry composition

There are 2 unique types of molecules in this entry. The entry contains 1281 atoms, of which 491 are hydrogens and 0 are deuteriums.

- Molecule 1 is a RNA chain called TAR RNA.

Mol	Chain	Residues	Atoms						Trace
			Total	C	H	N	O	P	
1	A	30	964	285	326	114	210	29	0

- Molecule 2 is a protein called Protein Tat.

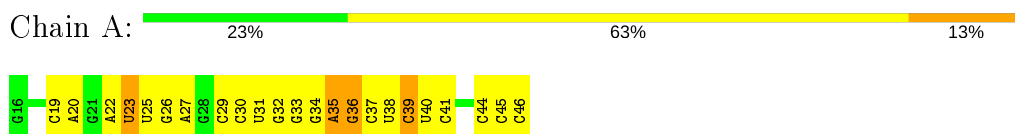
Mol	Chain	Residues	Atoms					Trace
			Total	C	H	N	O	
2	B	17	317	89	165	41	22	0

4 Residue-property plots [i](#)

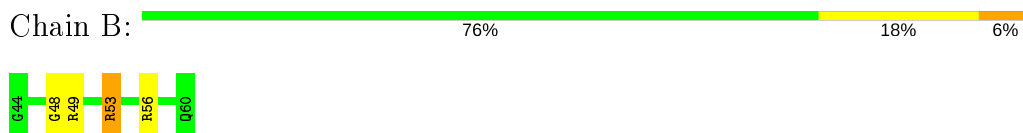
4.1 Average score per residue in the NMR ensemble

These plots are provided for all protein, RNA and DNA chains in the entry. The first graphic is the same as shown in the summary in section 1 of this report. The second graphic shows the sequence where residues are colour-coded according to the number of geometric quality criteria for which they contain at least one outlier: green = 0, yellow = 1, orange = 2 and red = 3 or more. Stretches of 2 or more consecutive residues without any outliers are shown as green connectors. Residues which are classified as ill-defined in the NMR ensemble, are shown in cyan with an underline colour-coded according to the previous scheme. Residues which were present in the experimental sample, but not modelled in the final structure are shown in grey.

- Molecule 1: TAR RNA



- Molecule 2: Protein Tat

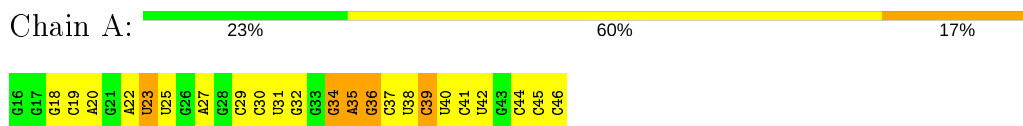


4.2 Scores per residue for each member of the ensemble

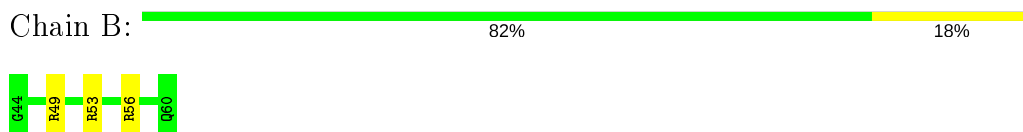
Colouring as in section 4.1 above.

4.2.1 Score per residue for model 1

- Molecule 1: TAR RNA



- Molecule 2: Protein Tat

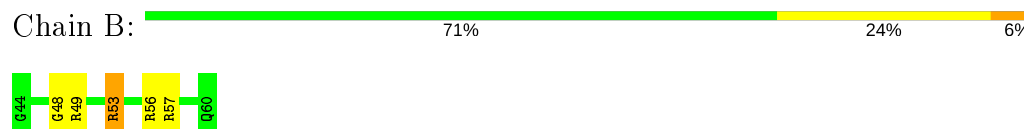


4.2.2 Score per residue for model 2

- Molecule 1: TAR RNA



- Molecule 2: Protein Tat

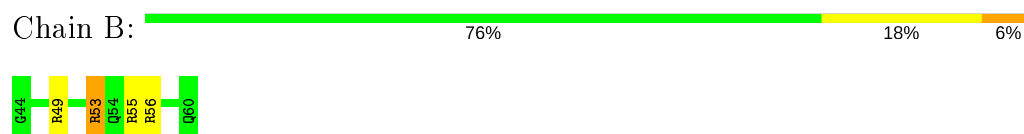


4.2.3 Score per residue for model 3

- Molecule 1: TAR RNA



- Molecule 2: Protein Tat

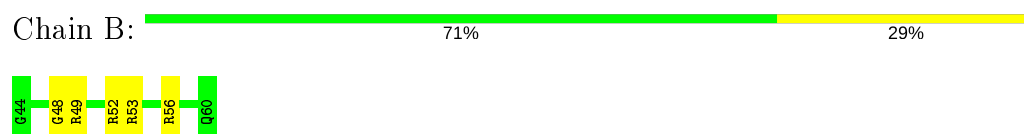


4.2.4 Score per residue for model 4

- Molecule 1: TAR RNA

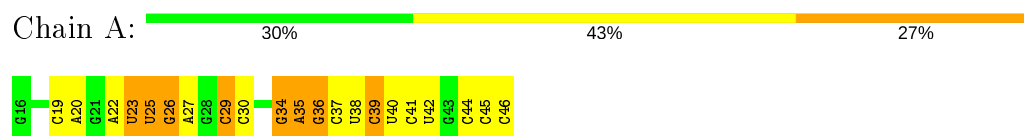


- Molecule 2: Protein Tat



4.2.5 Score per residue for model 5

- Molecule 1: TAR RNA

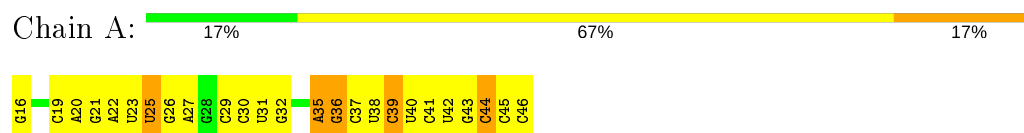


- Molecule 2: Protein Tat

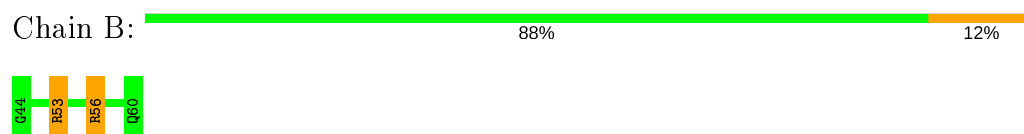


4.2.6 Score per residue for model 6

- Molecule 1: TAR RNA

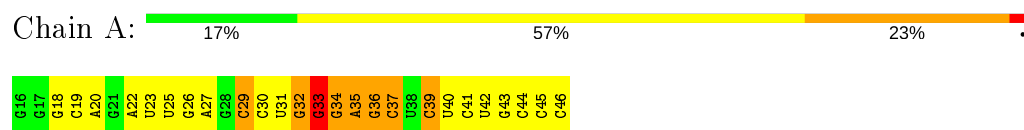


- Molecule 2: Protein Tat

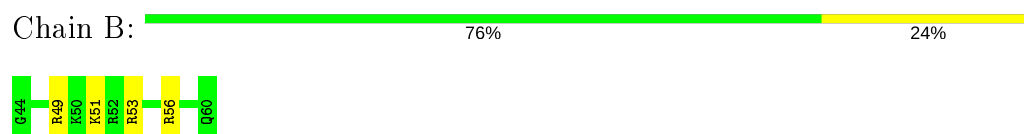


4.2.7 Score per residue for model 7

- Molecule 1: TAR RNA

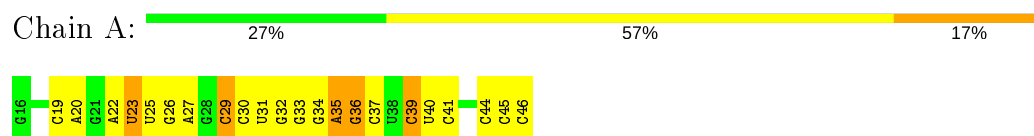


- Molecule 2: Protein Tat

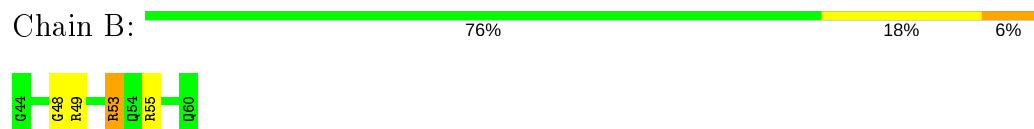


4.2.8 Score per residue for model 8

- Molecule 1: TAR RNA

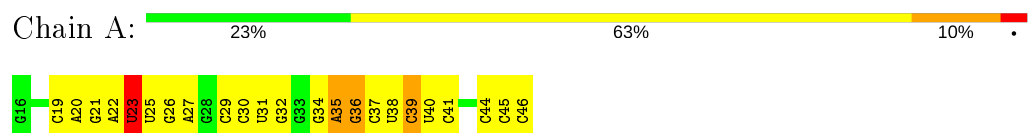


- Molecule 2: Protein Tat

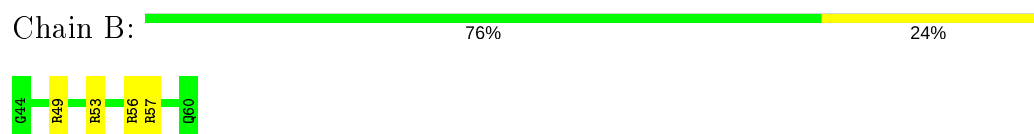


4.2.9 Score per residue for model 9

- Molecule 1: TAR RNA

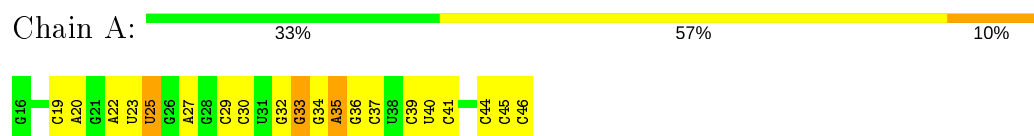


- Molecule 2: Protein Tat

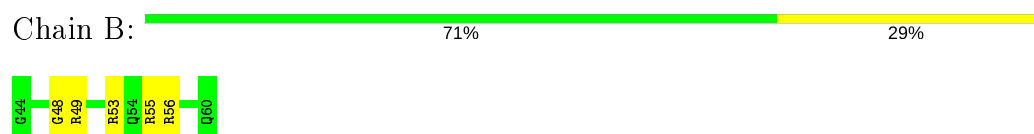


4.2.10 Score per residue for model 10

- Molecule 1: TAR RNA



- Molecule 2: Protein Tat



5 Refinement protocol and experimental data overview (i)

The models were refined using the following method: *simulated annealing*.

Of the 20 calculated structures, 10 were deposited, based on the following criterion: *structures with acceptable covalent geometry*.

The following table shows the software used for structure solution, optimisation and refinement.

Software name	Classification	Version
CYANA	structure calculation	
Amber	refinement	

The following table shows chemical shift validation statistics as aggregates over all chemical shift files. Detailed validation can be found in section 6 of this report.

Chemical shift file(s)	input_cs.cif
Number of chemical shift lists	1
Total number of shifts	447
Number of shifts mapped to atoms	447
Number of unparsed shifts	0
Number of shifts with mapping errors	0
Number of shifts with mapping warnings	0
Assignment completeness (well-defined parts)	51%

No validations of the models with respect to experimental NMR restraints is performed at this time.

COVALENT-GEOMETRY INFOmissingINFO

5.1 Too-close contacts (i)

In the following table, the Non-H and H(model) columns list the number of non-hydrogen atoms and hydrogen atoms in each chain respectively. The H(added) column lists the number of hydrogen atoms added and optimized by MolProbity. The Clashes column lists the number of clashes averaged over the ensemble.

Mol	Chain	Non-H	H(model)	H(added)	Clashes
All	All	7900	4910	4880	-

The all-atom clashscore is defined as the number of clashes found per 1000 atoms (including hydrogen atoms). The all-atom clashscore for this structure is -.

There are no clashes.

5.2 Torsion angles [i](#)

5.2.1 Protein backbone [i](#)

In the following table, the Percentiles column shows the percent Ramachandran outliers of the chain as a percentile score with respect to all PDB entries followed by that with respect to all NMR entries. The Analysed column shows the number of residues for which the backbone conformation was analysed and the total number of residues.

Mol	Chain	Analysed	Favoured	Allowed	Outliers	Percentiles	
2	B	15/17 (88%)	13±1 (84±5%)	2±1 (11±4%)	1±0 (5±3%)	4	27
All	All	150/170 (88%)	126 (84%)	17 (11%)	7 (5%)	4	27

All 3 unique Ramachandran outliers are listed below. They are sorted by the frequency of occurrence in the ensemble.

Mol	Chain	Res	Type	Models (Total)
2	B	48	GLY	5
2	B	56	ARG	1
2	B	51	LYS	1

5.2.2 Protein sidechains [i](#)

In the following table, the Percentiles column shows the percent sidechain outliers of the chain as a percentile score with respect to all PDB entries followed by that with respect to all NMR entries. The Analysed column shows the number of residues for which the sidechain conformation was analysed and the total number of residues.

Mol	Chain	Analysed	Rotameric	Outliers	Percentiles	
2	B	14/14 (100%)	13±1 (96±5%)	1±1 (4±5%)	33	81
All	All	140/140 (100%)	134 (96%)	6 (4%)	33	81

All 2 unique residues with a non-rotameric sidechain are listed below. They are sorted by the frequency of occurrence in the ensemble.

Mol	Chain	Res	Type	Models (Total)
2	B	53	ARG	5
2	B	56	ARG	1

5.2.3 RNA [i](#)

Mol	Chain	Analysed	Backbone Outliers	Pucker Outliers	Suiteness
1	A	29/30 (97%)	6±1 (20±3%)	2±1 (8±3%)	0.49±0.02
All	All	290/300 (97%)	58 (20%)	24 (8%)	0.49

The overall RNA backbone suiteness is 0.49.

All unique RNA backbone outliers are listed below:

Mol	Chain	Res	Type	Models (Total)
1	A	25	U	10
1	A	23	U	10
1	A	35	A	10
1	A	32	G	9
1	A	34	G	7
1	A	36	G	7
1	A	33	G	3
1	A	26	G	2

All unique RNA pucker outliers are listed below:

Mol	Chain	Res	Type	Models (Total)
1	A	39	C	9
1	A	25	U	7
1	A	35	A	3
1	A	33	G	3
1	A	32	G	2

5.3 Non-standard residues in protein, DNA, RNA chains [i](#)

There are no non-standard protein/DNA/RNA residues in this entry.

5.4 Carbohydrates [i](#)

There are no carbohydrates in this entry.

5.5 Ligand geometry [i](#)

There are no ligands in this entry.

5.6 Other polymers [i](#)

There are no such molecules in this entry.

5.7 Polymer linkage issues

There are no chain breaks in this entry.

6 Chemical shift validation [i](#)

The completeness of assignment taking into account all chemical shift lists is 51% for the well-defined parts and 51% for the entire structure.

6.1 Chemical shift list 1

File name: `input_cs.cif`

Chemical shift list name: `starch_output`

6.1.1 Bookkeeping [i](#)

The following table shows the results of parsing the chemical shift list and reports the number of nuclei with statistically unusual chemical shifts.

Total number of shifts	447
Number of shifts mapped to atoms	447
Number of unparsed shifts	0
Number of shifts with mapping errors	0
Number of shifts with mapping warnings	0
Number of shift outliers (ShiftChecker)	0

6.1.2 Chemical shift referencing [i](#)

No chemical shift referencing corrections were calculated (not enough data).

6.1.3 Completeness of resonance assignments [i](#)

The following table shows the completeness of the chemical shift assignments for the well-defined regions of the structure. The overall completeness is 51%, i.e. 429 atoms were assigned a chemical shift out of a possible 843. 0 out of 0 assigned methyl groups (LEU and VAL) were assigned stereospecifically.

	Total	¹ H	¹³ C	¹⁵ N
Backbone	39/85 (46%)	30/34 (88%)	9/34 (26%)	0/17 (0%)
Sidechain	98/174 (56%)	69/106 (65%)	29/46 (63%)	0/22 (0%)
Aromatic	6/15 (40%)	6/8 (75%)	0/6 (0%)	0/1 (0%)
Overall	429/843 (51%)	295/477 (62%)	114/285 (40%)	20/81 (25%)

The following table shows the completeness of the chemical shift assignments for the full structure. The overall completeness is 51%, i.e. 429 atoms were assigned a chemical shift out of a possible 843. 0 out of 0 assigned methyl groups (LEU and VAL) were assigned stereospecifically.

	Total	¹H	¹³C	¹⁵N
Backbone	39/85 (46%)	30/34 (88%)	9/34 (26%)	0/17 (0%)
Sidechain	98/174 (56%)	69/106 (65%)	29/46 (63%)	0/22 (0%)
Aromatic	6/15 (40%)	6/8 (75%)	0/6 (0%)	0/1 (0%)
Overall	429/843 (51%)	295/477 (62%)	114/285 (40%)	20/81 (25%)

6.1.4 Statistically unusual chemical shifts [i](#)

There are no statistically unusual chemical shifts.

6.1.5 Random Coil Index (RCI) plots [i](#)

The image below reports *random coil index* values for the protein chains in the structure. The height of each bar gives a probability of a given residue to be disordered, as predicted from the available chemical shifts and the amino acid sequence. A value above 0.2 is an indication of significant predicted disorder. The colour of the bar shows whether the residue is in the well-defined core (black) or in the ill-defined residue ranges (cyan), as described in section 2 on ensemble composition.

Random coil index (RCI) for chain B:

